Optical microscopy for nanoscale imaging : STED microscopy

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### Far field microscopy

- □ Nanoscale imaging in near field microscopes
  - For surface study only
  - Vulnerability to artifacts
  - Not suitable for soft materials
- Study of many natural processes requires knowledge about 3D volume of the sample
- Far field microscope can form a 3D image of a 3D object
  - Imaging deep into a biological sample
  - Non-invasive imaging



- $\hfill \hfill \hfill$
- **D** The illumination volume depends on  $\lambda$ , focal length and diameter of the illumination lens
- A point object is imaged into a diffraction limited volume in the image space
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#### Resolution of a microscope



point object and the corresponding image

two object points in the lateral direction whose images are just resolved two object points are indistinguishable in the image

two object points in the axial direction whose images are just resolved

Resolution: minimum separation between two point objects whose images are just resolved

Contributions from diffractions due to the illumination and detection lenses

□ Axial resolution is worse than lateral resolution

## Optical sectioning with a confocal microscope



- Confocal arrangement of focal point and pinhole blocks light from out of focus planes or points away from the optic axis
- □ The detector receives light mostly from the focal point
  - Image, free of out of focus blur, of a point object located at the focal point

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# Optical sectioning with a confocal microscope





Wide field image Confocal image (Source :www.olympusfluoview.com)

- Either the sample holding stage or the illumination spot is scanned
  - Scanning is controlled by a PC
- For each object point at the illumination spot, the detector signal is stored in the PC
- Results in an optically sectioned image (image corresponds to a sharply defined object plane, devoid of out of focus blur) of the sample
- Much better axial and marginally better lateral resolutions than a conventional (wide field) microscope
- □ Best resolution: lateral= $\sim\lambda/2$ , axial= $\sim\lambda$

## A beam scanning confocal microscope setup



#### Confocal fluorescence microscope



- □ Molecules (fluorophores) are excited with a laser beam of wavelength  $(\lambda_{EXC})$ , which than undergo a series of spontaneous emissions called fluorescence at the mean wavelength  $(\lambda_{EM})$
- $\hfill\square$  DBS: reflects  $\lambda_{EXC}$  and transmits  $\lambda_{EM}$
- $\hfill \hfill \hfill$

#### **Confocal fluorescence imaging**

- The target molecules are tagged with fluorescent probes or fluorophores
- Confocal detection of the fluorescent light in a beam scanning or stage scanning set up
- Fluorescence image provides information about the physical and



Confocal fluorescence image of human T cells (tagged with di-4-ANEPPDHQ fluorophores)

- chemical environment and orientation of the fluorophores and hence of the attached target molecules
- Best resolution working in the UV-visible range (lateral >200 nm, axial >500 nm)
  - Not enough for visualising light-matter interaction at nanoscale

#### Stimulated emission depletion (STED)



- **L**aser beam ( $\lambda_{EXC}$ ) excites a molecule to the upper electronic state
- $\hfill \label{eq:laserbeam}$  Another laser beam, called STED beam, at  $(\lambda_{\text{STED}})$  shines on the excited molecule
  - Stimulates it to undergo emission at ( $\lambda_{STED}$ )
  - No emission at ( $\lambda_{EM}$ ) i.e. No fluorescence from the excited molecule

#### **Resolution enhancement with STED**



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#### Nanometer resolution with saturated depletion Intensity level of the STED beam, indicated by vertical position



Fluorescence active volume

Normal power STED beam high power STED beam (GW/cm<sup>2</sup>)

- For normal STED beam intensity : only marginal improvement in resolution
- Depletion, and hence fluorescence intensity, has a nonlinear dependence on STED beam intensity
- Increasing the power of the STED beam to GW/cm<sup>2</sup> saturates depletion everywhere except along the optic axis
  - Lateral resolution ~28 nm

#### STED in a confocal fluorescence microscope



- Both excitation and STED beams are pulses following one another, usually derived from the same femto second laser
- Image is formed by scanning the stage or by scanning the beams

### Types of STED beams



- STED beams are generated using fixed phase plates or programmable diffractive elements such as a spatial light modulator
- □ Lateral resolution achieved ~28 nm
- Axial resolution achieved ~143 nm

#### Nanoscale imaging of fluorescent beads

#### Confocal image



#### STED image



XY plane images of 200 nm fluorescent beads (source: PhD thesis, B R Boruah, Imperial College London)

In biological science

Confocal image

STED image

QuickTime<sup>™</sup> and a TIFF (LZW) decompressor are needed to see this picture.

 Reveals nanopattern in the in SNAP-25 protein found in the plasma membrane of mamalian cells (source: Briefings in functional genomics and proteomics, Vol 5, No 4, 289-301)



(source: PNAS, 97, 15, 2000, 8206-8210)



 Revealing the structure of 3D colloidal crystals (source: Nano Letters, 8, 1309, 2008)

### Conclusion

- Near field microscopy can provide nanoscale resolution, however useful for surface study only
- Far field microscopy can be used to image 3D and biological samples
- Laser scanning confocal microscope is a far field microscopy technique that can provide optically sectioned image
  - Can image either with reflected or fluorescence light
  - Useful for 3D visualisation of the sample
  - Resolution is limited by the diffraction effects
- Stimulated emission depletion phenomenon can be used in a confocal fluorescence microscope to get resolution beyond the diffraction barrier
- STED microscope has been used in the study of nano structures in living and non-living samples